## Supplementary information

## **Supplementary methods**

# **Plasmid construction**

Streptavidin refers to core streptavidin (Sano et al, 1995) with His<sub>6</sub> at the C-terminus expressed from pET21a(+) (Howarth et al, 2006). The strong mutant was generated by introducing the S52G R53S mutation via QuikChange<sup>TM</sup> (Stratagene) using the its complement: 5'following primer and reverse GTTGGTAACGCTGAAGGTAGCTACGTTCTGACCGGTCG. A86D streptavidin 5'was generated in the same way with the primer GAAAAACAACTACCGTAACGATCACTCCGCTACCAC and its reverse complement. Then the weak mutant, A86D H87G, was generated from A86D streptavidin with the primer 5'- CTACCGTAACGATGGCTCCGCTACCACCTGG and its reverse complement. Such mutants with suitable off-rates were found via combined random and rational mutagenesis, based on previous mutations and screens of streptavidin (Aslan et al, 2005; Laitinen et al, 2006; Levy and Ellington, 2008). The mutations were confirmed by DNA sequencing.

## Streptavidin expression and purification

Streptavidin and its mutants were expressed as described (Howarth and Ting, 2008). Briefly, *E. coli* BL21 (DE3) RIPL (Stratagene) was grown to  $OD_{600}$  0.9 at 37 °C, induced with IPTG, and incubated for a further 4 hr at 37 °C. Inclusion bodies were isolated, dissolved in guanidinium hydrochloride, and refolded by rapid dilution into PBS. Soluble protein was then further purified by Ni-NTA and dialyzed into PBS. All versions of streptavidin showed good solubility and were entirely tetrameric (data not shown).

# **Off-rate** assay

The off-rate of biotin-4-fluorescein (B4F) from streptavidin or its strong or weak mutants was measured as described (Howarth *et al*, 2006). The binding of B4F to an excess of binding protein quenches fluorescein emission (Kada *et al*, 1999). As B4F dissociates at 37 °C in the presence of excess free biotin, the fluorescence of B4F recovers.

#### **Supplementary references**

- Aslan FM, Yu Y, Mohr SC, and Cantor CR (2005) Engineered single-chain dimeric streptavidins with an unexpected strong preference for biotin-4-fluorescein. *Proc Natl Acad Sci USA* **102:** 8507-8512
- Howarth M, Chinnapen DJ, Gerrow K, Dorrestein PC, Grandy MR, Kelleher NL, El Husseini A, Ting AY (2006) A monovalent streptavidin with a single femtomolar biotin binding site. *Nat Methods* 3: 267-273
- Howarth M and Ting AY (2008) Imaging proteins in live mammalian cells with biotin ligase and monovalent streptavidin. *Nat Protoc* **3:** 534-545
- Kada G, Falk H, Gruber HJ (1999) Accurate measurement of avidin and streptavidin in crude biofluids with a new, optimized biotin-fluorescein conjugate. *Biochim Biophys Acta* 1427: 33-43
- Laitinen OH, Hytonen VP, Nordlund HR, and Kulomaa MS (2006) Genetically engineered avidins and streptavidins. *Cell Mol Life Sci* 63: 2992-3017
- Levy M, Ellington AD (2008) Directed evolution of streptavidin variants using in vitro compartmentalization. *Chem Biol* **15**: 979-989
- Sano T, Pandori MW, Chen X, Smith CL, Cantor CR (1995) Recombinant core streptavidins. A minimum-sized core streptavidin has enhanced structural stability and higher accessibility to biotinylated macromolecules. *J.Biol.Chem* **270**: 28204-28209

#### **Supplementary legends**

## Supplementary Figure 1

Changed biotin-conjugate off-rate for weak and strong mutants of streptavidin. Wildtype streptavidin and weak (A86D H87G) or strong (S52G R53S) mutants were incubated with biotin-4-fluorescein, quenching its fluorescence. On addition of excess free biotin, biotin-4-fluorescein dissociated and the increase in fluorescence was observed at 37 °C and pH 7.4. Mean of duplicate (weak mutant) or triplicate (WT and strong mutant) measurements  $\pm 1$  s.d.

